MECHANISMS OF DISEASE

Pemphigus, Bullous Impetigo, and the Staphylococcal Scalded-Skin Syndrome

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Pemphigus, which is caused by autoantibodies, and bullous impetigo (including its generalized form, the staphylococcal scalded-skin syndrome), which is caused by *Staphylococcus aureus*, are seemingly unrelated diseases. However, 200 years ago, astute clinicians realized that these diseases had enough clinical similarities to call bullous impetigo and the scalded-skin syndrome in infants “pemphigus neonatorum.” In this review we explain how a common mechanism accounts for the clinical overlap of these blistering diseases of the skin, and how the unraveling of the molecular pathophysiology of pemphigus provided the clues that were necessary to determine the mechanism of the formation of blisters in bullous impetigo and the staphylococcal scalded-skin syndrome. We also discuss how this new understanding of the pathophysiology of pemphigus could improve the diagnosis and treatment of this potentially life-threatening disease.

Pemphigus

There are two major types of pemphigus, pemphigus vulgaris and pemphigus foliaceus.® Patients with pemphigus vulgaris present with blisters and erosions of mucous membranes and skin. There are two subtypes of pemphigus vulgaris: the mucosal-dominant type, with mucosal lesions but minimal skin involvement, and the mucocutaneous type, with extensive skin blisters and erosions in addition to mucosal involvement (Fig. 1A). Patients with pemphigus foliaceus have scaly and crusted superficial erosions of the skin but not of mucous membranes (Fig. 1B).

The blisters of pemphigus vulgaris are characterized by a loss of cell adhesion in the deep epidermis, just above the basal layer (Fig. 1C), whereas in pemphigus foliaceus, the loss of cell adhesion is in the more superficial epidermis, just below the stratum corneum, which is the layer of dead keratinocytes that forms the barrier of the skin (Fig. 1D).

The blood of patients with pemphigus contains IgG antibodies that bind to the surface of normal keratinocytes; this binding is shown with the use of immunofluorescence (Fig. 1E and 1F). Immunofluorescence staining also shows IgG antibodies on the surface of the keratinocytes in biopsy specimens of the skin from patients with pemphigus.

These antibodies, which are autoantibodies because they react with the patient’s own cells, are directly pathogenic — that is, they can cause loss of adhesion between keratinocytes, which results in blistering. When injected into neonatal mice, human IgG from patients with pemphigus vulgaris or pemphigus foliaceus binds to the surface of the epidermal keratinocytes (Fig. 1I) and causes blisters (Fig. 1G) with the typical histologic features of pemphigus vulgaris (Fig. 1H) or pemphigus foliaceus (Fig. 2D).4,5

Therefore, pemphigus vulgaris and pemphigus foliaceus are related in that they
are blistering diseases caused by autoantibodies that interfere with adhesion between keratinocytes. They can be differentiated clinically by the presence or absence of involvement of the mucous membranes, and histologically by the epidermal level where the blistering occurs.

DESCMOGENS

Because keratinocyte adhesion is defective in patients with pemphigus foliaceus and pemphigus vulgaris, it was logical to wonder whether the autoantibodies in patients with these disorders bind adhesion molecules in desmosomes. Desmosomes are cell adhesion structures that are especially prominent in the epidermis and mucous membranes. Transmembrane molecules that were discovered in desmosomes belong to two families of proteins — desmogleins and desmocollins (Fig. 3). Both families are related to cadherins, which are known to modulate cell–cell adhesion. For this reason, desmogleins and desmocollins are now called desmosomal cadherins, which presumably modulate adhesion within desmosomes.

Serum from patients with pemphigus foliaceus contains antibodies that bind to desmoglein 1, a member of the desmoglein family. The antigen recognized by the autoantibodies in patients with pemphigus vulgaris was cloned by using immunostaining with the autoantibodies to identify clones from a phage library that expressed proteins from normal keratinocytes. This screening showed that the pemphigus vulgaris antigen was a previously unknown member of the desmoglein family. The antigen was subsequently called desmoglein 3. Thus, pemphigus vulgaris and pemphigus foliaceus, which are related but distinct diseases, were found to have related but distinct autoantigens — desmoglein 3 and desmoglein 1, respectively.

PATHOGENICITY OF ANTIDESMOGEN Antibodies

Antidesmoglein antibodies cause blisters. The injection of affinity-purified autoantibodies against desmoglein 1 or desmoglein 3 into mice causes blisters with the typical histologic features of pemphigus foliaceus or pemphigus vulgaris, respectively. Furthermore, the gross and microscopic lesions of pemphigus vulgaris can be induced in mice by intraperitoneal inoculation of hybridoma cells that produce monoclonal anti–desmoglein 3 antibodies. In addition, desmoglein 1 or desmoglein 3 can adsorb the corresponding pathogenic antibodies from pemphigus serum. When monitored in individual patients, the titers of anti–desmoglein 1 and anti–desmoglein 3 IgG autoantibodies in serum, as measured by indirect immunofluorescence or enzyme-linked immunosorbent assay (ELISA), generally correlate with disease activity.

Pemphigus serum may bind antigens other than desmoglein 1 and desmoglein 3, but the clinical significance of these other antibodies is not well understood. For example, a subgroup of IgG anti–desmoglein 1 autoantibodies cross-reacts with desmoglein 4, but these antibodies have no pathogenic effect. Antibodies in pemphigus serum can bind to other antigens, such as acetylcholine receptors, but they have not been shown to directly mediate blister formation. Nevertheless, acetylcholine receptors may modulate disease activity in some patients who have pemphigus vulgaris.

DESMOGEN COMPENSATION

Clinically different subgroups of patients with pemphigus have characteristic profiles of antidesmoglein antibodies (Table 1). These autoantibody patterns, and the distribution of desmoglein isoforms in the epidermis and mucous membranes, suggest that desmoglein compensation can explain blister localization in patients with pemphigus vulgaris and pemphigus foliaceus. The desmoglein compensation theory rests on the following two observations: anti–desmoglein 1 or anti–desmoglein 3 autoantibodies inactivate only the corresponding desmoglein, and functional desmoglein 1 or desmoglein 3 alone is usually sufficient for cell–cell adhesion.

Desmoglein compensation has been validated experimentally in the neonatal-mouse model of pemphigus. The injection of anti–desmoglein 1 autoantibodies into mice in which the desmoglein 3 (Dsg3) gene has been deleted causes blistering in areas that are protected from blistering by desmoglein 3 in normal mice. Conversely, transgenic mice that have been engineered to express desmoglein 3 in areas that normally express only desmoglein 1 are protected from the blistering caused by anti–desmoglein 1 antibodies. Finally, transgenic expression of desmoglein 1 in areas
that normally express only desmoglein 3 can at least partially correct defective cell–cell adhesion due to the genetic loss of desmoglein 3 in knockout mice.28 Because the distribution of desmogleins in neonatal skin is similar to the distribution of desmogleins in mucous membranes, which do not blister in pemphigus foliaceus (Fig. 4A), desmoglein compensation explains why blisters usually do not develop in neonates whose mothers have pemphigus foliaceus, even though maternal autoantibodies cross the placenta and bind to the neonatal epidermis.27

**Pemphigus Autoantibodies and Loss of Adhesion of Keratinocytes**

**Inactivation of Desmoglein**

Some investigators proposed that pemphigus autoantibodies act by initiating a proteolytic cascade that nonspecifically cleaves cell-surface molecules.29,30 Subsequent studies, however, did not support this hypothesis.31 Other investigations have shown that anti–desmoglein 3 and anti–desmoglein 1 antibodies inactivate their targeted desmogleins with considerable specificity. The lesions caused by these antibodies are remarkably similar, if not identical, to the lesions caused by direct inactivation of desmoglein 3 or desmoglein 1. For example, the pathological features of the skin of knockout mice with an inactivated Dsg3 gene are similar to those in patients with pemphigus vulgaris and to those in mice injected with anti–desmoglein 3 antibodies.32 And in both mice and humans, exfoliative toxin, which specifically cleaves desmoglein 1, causes blisters that are identical to the blisters caused by the anti–desmoglein 1 autoantibodies in cases of pemphigus foliaceus.33 These findings, together with the desmoglein compensation theory, suggest that pemphigus autoantibodies inactivate only their targeted desmoglein and do not cause a generalized loss of function of cell-surface adhesion molecules.

**Direct and Indirect Effects of Pemphigus Antibodies**

Whether pemphigus autoantibodies act directly or indirectly is controversial. There is evidence that pemphigus autoantibodies block adhesion by directly interfering with desmoglein transinteractions (i.e., interactions of desmoglein on one cell with itself or with desmocollins on the adjacent cell). Studies have shown that fragments of pemphigus autoantibodies that contain only a single antigen-binding domain and lack the effector (Fc) region of antibodies can induce blisters in mice14–16 and, because they lack the ability to cross-link cell-surface molecules, probably interfere directly with adhesion. Furthermore, a mouse monoclonal anti–desmoglein 3 IgG antibody that binds to the N-terminal adhesive interface of desmoglein 3 induces pemphigus vulgaris lesions in mice, whereas other monoclonal antibodies that react with functionally less important parts of desmoglein 3 do not cause lesions in mice.14

Alternatively, the results of a recent study using single-molecule atomic-force measurements, a biomechanical method that measures the degree of protein–protein binding, suggest that anti–desmoglein 1 IgG antibodies in pemphigus foliaceus serum do not interfere directly with desmoglein 1 adhesive transinteractions.37 In this extracellular system, the binding of desmoglein 1 to itself was not inhibited by pathogenic anti–desmoglein 1 antibodies. Other studies suggest that direct functional
inactivation of desmoglein may not be sufficient to cause the blisters and that pemphigus autoantibodies may act through a more complex signaling mechanism. The addition of IgG from pemphigus vulgaris serum to cultured keratinocytes induces a number of signals, including a transient increase in intracellular calcium and inositol 1,4,5-triphosphate, activation of protein kinase C, and phosphorylation of desmoglein 3, which in turn may lead to the internalization of desmoglein 3.
from cell surfaces, with resultant depletion of desmoglein 3 from desmosomes.

Pemphigus vulgaris IgG is also reported to induce activation of other signaling pathways that may result in reorganization of the cytoskeleton, apoptosis of keratinocytes, or both. Further studies are needed to clarify whether such signaling is involved in blister formation in vivo, because most of the studies on signal transduction were performed in vitro with the use of cultured keratinocytes.

**BULLOUS IMPETIGO AND THE STAPHYLOCOCCAL SCALDED-SKIN SYNDROME**

Pemphigus neonatorum

A condition of infants that resembled pemphigus foliaceus was called pemphigus neonatorum by clinicians in the 18th and 19th centuries. Pemphigus neonatorum was actually bullous impetigo or the staphylococcal scalded-skin syndrome. The observation of the similarities among pemphigus foliaceus, bullous impetigo, and the scalded-skin syndrome was the first suggestion that the pathophysiology of pemphigus might also apply to the other disorders.

**PRODUCTION OF EXFOLIATIVE TOXIN BY STAPHYLOCOCCUS**

Staphylococcal skin infections are among the most common skin diseases in children. Classic studies more than 30 years ago showed that the blisters in bullous impetigo and the scalded-skin syndrome are caused by exfoliative toxin released by staphylococcus. Subsequently, it was discovered that two major serotypes of this toxin, A and B, cause bullous impetigo and the scalded-skin syndrome. In patients with bullous impetigo, the toxin produces blisters locally at the site of infection, whereas in cases of the scalded-skin syndrome, it circulates throughout the body, causing blisters at sites distant from the infection. The risk of death from the scalded-skin syndrome is less than 5% among children, but among adults, the syndrome usually occurs in those who have renal disease or who are immunosuppressed, and the risk of death can be as high as 60%. Exfoliative toxin injected into neonatal mice causes the blisters typical of bullous impetigo and the scalded-skin syndrome. Exactly how exfoliative toxin causes these blisters was controversial for many years. However, when exfoliative toxin A and exfoliative toxin B were cloned, they were found to have amino acid sequences and crystal structures suggestive of serine proteases. In some crystal solutions this structure was slightly irregular in that one peptide bond in the catalytic site was rotated 180 degrees in the wrong direction to form an active catalytic pocket. These findings indicated that exfoliative toxin might not be active as a protease unless it bound a specific substrate that was able to reform its catalytic pocket.

The specific substrate for exfoliative toxin remained elusive for several years. However, four important clues from studies of pemphigus foliaceus indicated that the substrate might be desmoglein 1. First, the clinical features of the scalded-skin syndrome (and its localized form, bullous impetigo) (Fig. 2B) resemble those of pemphigus foliaceus (Fig. 2A). Second, in cases of the scalded-skin syndrome, the blisters caused by exfoliative
toxin appear only on the skin, not on mucous membranes (the same distribution as the lesions caused in pemphigus foliaceus). Third, blisters in bullous impetigo and the scalded-skin syndrome (Fig. 2C) are histologically indistinguishable from those in pemphigus foliaceus (Fig. 1D). Finally, the injection of pemphigus foliaceus IgG antibodies or exfoliative toxin into neonatal mice causes blisters with identical gross and histologic features (Fig. 2D and 2E). These clues suggested that exfoliative toxin, like pemphigus foliaceus IgG, might target desmoglein 1. If desmoglein 1 was specifically cleaved by exfoliative toxin, then — just as in pemphigus foliaceus — desmoglein compensation would account for the localization of the blisters in bullous impetigo and the scalded-skin syndrome only in the superficial epidermis, not in mucous membranes (Fig. 4B).

Indeed, studies show that exfoliative toxins do cleave desmoglein 1, but not the closely related desmoglein 3, on cultured human keratinocytes and keratinocytes in the skin of neonatal mice. Exfoliative toxin also cleaves recombinant desmoglein 1, but not desmoglein 3 or the intercellular adhesion molecule E-cadherin, in solution, showing a direct proteolytic effect of the toxin. Moreover, exfoliative toxin efficiently cleaves a particular peptide bond in desmoglein 1 at a calcium-binding site, and its ability to do so depends on the conformation of the site — that is, the toxin cannot cleave denatured desmoglein 1. The ability of the toxin to cleave desmoglein 1 also depends on amino acids about 100 residues upstream of the cleavage site.

These findings suggest that exfoliative toxin cleaves desmoglein 1 by the key-in-lock mechanism that is common to many proteolytic enzymes with limited substrate specificities. This remarkable mechanism efficiently targets one molecule — desmoglein 1 — that allows staphylococcus to grow below the epidermal barrier but superficially enough to be contagious by skin contact.

**DIAGNOSIS AND THERAPY**

**BULLOUS IMPETIGO AND THE STAPHYLOCOCCAL SCALDED-SKIN SYNDROME**

The staphylococcal scalded-skin syndrome can sometimes resemble another widespread blistering disease, toxic epidermal necrolysis, which is usually caused by a drug reaction. These two diseases can be differentiated quickly by examining a frozen section of a biopsy specimen, which shows a superficial epidermal blister in cases of the scalded-skin syndrome and a subepidermal blister with necrotic keratinocytes in toxic epidermal necrolysis. Treatment of patients with bullous impetigo...
The presence of IgG autoantibodies that bind to the keratinocyte surface or desmogleins is the gold standard for the diagnosis of pemphigus and its differentiation from other vesiculobullous or pustular diseases. Until recently, direct immunofluorescence (on the skin of patients) or indirect immunofluorescence (in serum from patients) was the standard method of detecting antibodies that bind to the surface of keratinocytes. However, ELISA with the use of recombinant desmoglein 1 and desmoglein 3 as antigens is much simpler and more quantifiable than immunofluorescence.

ELISA to identify antibodies against desmoglein 1 and desmoglein 3 can be used not only for diagnosing the type of pemphigus but also for distinguishing subtypes of pemphigus vulgaris (Table 1).

In general, pemphigus is treated by suppressing the immune system to blunt the autoimmune response. However, more targeted therapy may be possible. For example, recent case reports indicate that rituximab, an anti-CD20 monoclonal antibody that targets B cells (lymphocytes that mature to antibody-producing plasma cells), can be very effective in treating patients with pemphigus that is refractory to more standard immunosuppressive therapy.

It might be possible to focus the development of new, more specific treatments for pemphigus on the autoantibodies themselves, especially if the population of pathogenic autoantibodies in the disease is not diverse. The extent of their diversity has been studied by phage display, which allows the cloning of monoclonal antidesmoglein antibodies from patients with pemphigus. This method has been used in studies of one patient with pemphigus vulgaris and one with pemphigus foliaceus (unpublished data); the results indicate that the populations of antidesmoglein antibodies in both cases were highly restricted, as judged by the lack of diversity of heavy chains in populations of monoclonal antibodies. This finding contrasts with the usual population of antibacterial antibodies, which are constructed from a large array of heavy chains and light chains. The possibility of a common structural pattern among pemphigus autoantibodies could have clinical implications for targeted therapy against a subpopulation of antibodies, as opposed to the suppression of the general production of antibodies.

Finally, understanding how T and B cells contribute to the antidesmoglein response could provide another basis for therapy. A mouse model of pemphigus vulgaris was developed by the transfer of T cells and B cells from Dsg3−/− knockout mice, in which desmoglein 3 acts like a foreign antigen, into Rag2−/− immunodeficient mice that express desmoglein 3. The recipients of T cells and B cells from Dsg3−/− mice produced anti–desmoglein 3 antibodies, and pemphigus vulgaris lesions developed. T and B cells were necessary for both antibody production and blister formation. These results are consistent with increasing evidence of the role of autoreactive T cells in regulating the production of pathogenic IgG autoantibodies in humans. Furthermore, desmoglein 3–specific regulatory T cells were recently shown to be involved in the maintenance of peripheral tolerance to desmoglein 3 in healthy persons. Thus, manipulation of regulatory T cells that are actively engaged in the control of a variety of physiologic and pathologic immune responses may provide a promising option for the treatment of pemphigus.

**Table 1. Antidesmoglein Antibody Profiles in Subtypes of Pemphigus.**

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<thead>
<tr>
<th>Subtype</th>
<th>IgG Autoantibody</th>
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<tr>
<td></td>
<td>Anti–Desmoglein 1</td>
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<tr>
<td>Pemphigus foliaceus</td>
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</tr>
<tr>
<td>Pemphigus vulgaris</td>
<td>No</td>
</tr>
<tr>
<td>Mucocutaneous</td>
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**CONCLUSIONS**

Further studies are needed to refine our understanding of the pathogenic mechanisms of pemphigus. This knowledge will provide the foundation for developing more targeted and better...
Figure 4. The Desmoglein Compensation Theory and the Sites of Blister Formation in Pemphigus, Bullous Impetigo, and the Staphylococcal Scalded-Skin Syndrome.

The colored triangles and rectangles represent the distribution of Dsg1 and Dsg3 in the skin and mucous membranes. Anti-Dsg1 IgG autoantibodies in serum from patients with pemphigus foliaceus cause superficial blisters in the skin; no blisters form in the lower epidermis or mucous membranes, because Dsg3 maintains cell–cell adhesion in those areas (Panel A). In bullous impetigo and the staphylococcal scalded-skin syndrome, exfoliative toxin (ET) produced by S. aureus acts like Dsg1-specific molecular scissors and exclusively cleaves Dsg1 but not Dsg3, resulting in only superficial epidermal blisters, because Dsg3 compensates in other areas (Panel B). Serum from patients with mucosal-dominant pemphigus vulgaris contains only anti-Dsg3 IgG, which causes mucosal blisters and erosions where there is no significant compensation by Dsg1 (Panel C).
tolerated therapies for these disfiguring and life-threatening diseases.

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